



Figure 1. Chest X-ray showing heterogenous opacity involving right middle and lower zones suggestive of ARDS.

virological study by real time polymerase chain reaction (RT-PCR) (on day 8 of illness). His urine tested weakly positive for NiV, while throat swab and blood were negative. Repeat blood culture samples were again sterile. Antiviral was changed to oral ribavirin (2-g loading dose followed by 4 g/day in divided doses).

On day 8, his blood pressure rose to 200/120mm Hg. Electrocardiogram showed sinus tachycardia and echocardiogram was normal. On day 9, he developed hypotension requiring inotropic support (with dopamine and noradrenaline). Repeat blood, urine, and throat swab samples (sent 4 days after the initial sample) tested strongly positive for NiV. He expired on day 14 (i.e., 17th day of illness). The timeline of events have been outlined in Table 1.

Discussion

NiV belongs to Henipavirus genus of Paramyxoviridae family. Pteropus bats, popularly known as flying foxes or fruit bats, are the natural reservoirs of the virus. The first incidence of the NiV was reported simultaneously in pigs and humans in Malaysia in years 1998–1999 [1]. The name was derived from Kampung Sungai Nipah or Nipah River Village, where the virus was first isolated. Pigs were the intermediate hosts displaying symptoms of infectious respiratory and neurologic disease [2].

The changes in ecological conditions, in the form of urbanization and deforestation, have led to the emergence of NiV into the pig population and subsequently into the human population. The establishment of pig farms in Malaysia within the territory of the natural host led to the introduction of the virus into the pig population; with subsequent spread in southern Malaysia and Singapore. The direct contact with pigs (which act as amplifying host) or fresh pig products was responsible for the transmission of the virus to humans [3].

In Philippines, the virus to human transmission was thought to be due to direct exposure to infected horses, consumption of undercooked meat of infected horses, or

from contact with contaminated body fluids while slaughtering the infected horses [4].

The outbreaks in Bangladesh and India were believed to be either due to direct contact with bats or by contact with material contaminated by them like date palm sap. The Bangladesh outbreaks have identified different routes of transmission like climbing trees (probably contaminated with infected date palm sap), contact with sick persons, and contact with sick animals.

The NiV infection is characterized by severe, rapidly progressive encephalitis, carrying a high mortality rate [5]. The symptoms usually occur within 3 to 14 days of exposure, with an incubation period ranging from 5 to 14 days. However, an incubation period as long as 60 days has been reported [6]. Malaysia and Singapore witnessed severe febrile encephalitic disease [7]. Fever, headache, dizziness, and vomiting were the main symptoms. More than 50% of the patients had decreased consciousness and brainstem dysfunction [5]. The same was the situation in Bangladesh resulting in very high case fatalities [8]. In India, there were two outbreaks. The first occurred at Siliguri, West Bengal during the year 2001, affecting 66 people with acute encephalitis and resulting in 45 deaths. Fever, headache, vomiting, altered sensorium, myalgia, respiratory distress, and involuntary movements or convulsions were observed. Patients were normotensive on admission and later developed hypertension prior to death [9]. The second outbreak took place in Nadia district of West Bengal. Fever, headache, and myalgia were the main presenting complaints. Some also had episodes of vomiting, disorientation, respiratory distress [10]. In Philippines, majority patients presented with encephalitis, while others had influenza like symptoms and meningitis [4].

The diagnostic tests include virus isolation, nucleic acid amplification tests, and serology [11]. Virus isolation and RT-PCR from throat and nasal swabs, cerebrospinal fluid, blood, and urine should be performed during the early course of the disease. In later stages, antibody detection by ELISA (IgM and IgG) can be used. In fatal cases, immunohistochemistry on tissues collected during autopsy should be performed to confirm the diagnosis. The standard test for detection of anti-NiV antibody is serum neutralization test [12].

The treatment is mainly supportive care; with no medications or vaccines available as of now. Ribavirin, a broad spectrum antiviral, can cross the blood brain barrier and may be useful to reduce the mortality in cases of acute NiV encephalitis and the duration of ventilator support [13]. m102.4, a neutralizing human mechanical antibody, that recognize the receptor binding domain of the NiV G glycoprotein has been successfully tested in animal model [14]. Favipiravir, a purine analogue that inhibit viral RNA-dependent RNA polymerase, shows promising results in NiV infection [15]. Various studies have shown that balapiravir may be active against NiV.

Table 1. Timeline of events.

DAY	EVENT
0	Fever, watery loose stools and non-projectile vomiting, throat pain
3	Presented to hospital
6	Developed binocular horizontal diplopia on gaze to right
7	Gait ataxia, segmental sweating over upper trunk
8	ARDS – Intubated and put on mechanical ventilation Urine for NiV (RT-PCR) – Weakly Positive Blood and throat swab specimen for NiV (RT-PCR) – Negative Oral ribavirin started
11	Accelerated hypertension
12	Repeat Urine, blood and throat swab specimen for NiV (RT-PCR) – Strongly Positive
13	Hypotension requiring inotropic support
17	Expired

The risk of bat to human transmission can be reduced by decreasing bat access to date palm sap and by boiling freshly collected date palm juice. Bat bitten fruits should be avoided. All fruits should be washed thoroughly and the skin should be peeled before eating. The risk of human-to-human transmission can be reduced by avoiding close contact with infected patients. Health care workers and those taking care of infected individuals should use gloves and personal protective equipment. Regular hand washing is mandatory. Disinfection of the equipment and environment, and proper waste management is essential in preventing transmission. Protective clothing and gloves should be worn while handling sick animal and their tissue in order to reduce the risk of animal to human transmission.

The patient being reported was our first experience with NiV. Unlike the usual findings of leucopenia and thrombocytopenia, our patient had a normal complete blood count. Cranial nerve paralysis, especially ptosis, brain stem dysfunction evidenced by hypertension, tachycardia, segmental sweating, and nystagmus were observed in the Malaysian outbreak; but cerebellar dysfunction was an unusual manifestation which was seen in our case. The patient described was one of the initial cases which marked the beginning of the NiV outbreak of May 2018 in the “Malabar” region of Kerala state in India. However, early diagnosis, robust infection control measures along with adequate and proper barrier methods helped to curtail the outbreak rapidly.

Conclusion

NiV in humans presents with a wide spectrum of clinical manifestations, ranging from asymptomatic infection to acute respiratory infection and fatal encephalitis. Fruit bats are the natural reservoirs of NiV. The virus can be transmitted to humans from animals (such as bats or pigs), or contaminated foods. Human-to-human transmission is also possible. The primary treatment for humans is supportive care. There is no treatment or vaccine available for either people or animals. Ribavirin may be useful in acute NiV encephalitis.

List of Abbreviations

NiV	Nipah virus
VCA	Viral Capsid Antigen
ARDS	acute respiratory distress syndrome
RT-PCR	real time polymerase chain reaction

Consent for publication

Obtained from Wife as patient expired.

Ethical approval

Yes (since this is not a research work, only verbal approval was obtained with importance to patient bystander consent).

Author details

Bhargavan Pallivalappil¹, Ummer Karadan², Jayakrishnan Chellenton³, Robin George Manappallil^{4*}

1. Senior Consultant, Department of Internal Medicine, Baby Memorial Hospital, Calicut, Kerala, India
2. Senior Consultant, Department of Neurology, Baby Memorial Hospital, Calicut, Kerala, India
3. Consultant, Department of Neurology, Baby Memorial Hospital, Calicut, Kerala, India
4. Consultant, Department of Internal Medicine, Baby Memorial Hospital, Calicut, Kerala, India

References

1. MMWR. Outbreak of Hendra-like virus-Malaysia & Singapore 1998–1999. *Morb Mortal Wkly Rep.* 1999;48:265–9.
2. Chua KB, Bellini WJ, Rota PA, Harcourt BH, Tamin A, Lam SK, et al. Nipah virus: a recently emergent deadly paramyxovirus. *Science.* 2000;288:1432–5. <https://doi.org/10.1126/science.288.5470.1432>
3. Chua KB, Goh KJ, Wong KT, Kamarulzaman A, Tan PS, Ksiazek TG, et al. Fatal encephalitis due to Nipah virus among pig farmers in Malaysia. *Lancet.* 1999;354:1257–9. [https://doi.org/10.1016/S0140-6736\(99\)04299-3](https://doi.org/10.1016/S0140-6736(99)04299-3)
4. Ching PK, de los Reyes VC, Sucaldito MN, Tayag E, Columa-Vingno AB, Malbas FF, et al. Outbreak of henipavirus infection, Philippines, 2014. *Emerg Infect Dis.* 2015;21:328–31. <https://doi.org/10.3201/eid2102.141433>
5. Goh KJ, Tan CT, Chew NK, Tan PS, Kamarulzaman A, Sarji SA, et al. Clinical features of Nipah virus encephalitis among pig farmers in Malaysia. *N Engl J Med.* 2000;342:1229–35. <https://doi.org/10.1056/NEJM200004273421701>
6. Tan KS, Tan CT, Goh KJ. Epidemiological aspects of Nipah virus infection. *Neurol J SE Asia.* 1999;4:77–81.

7. Parashar UD, Sunn LM, Ong F, Mounts AW, Arif MT, Ksiazek TG, et al. Case-control study of risk factors for human infection with a new zoonotic paramyxovirus, Nipah virus in Malaysia. *J Infect Dis.* 2000;181:1755–9. <https://doi.org/10.1086/315457>
8. International Centre for Diarrheal Disease Research, Bangladesh (ICDDR). Outbreaks of encephalitis due to Nipah/Hendra-like viruses. Western Bangladesh. *Health Sci Bull.* 2003;1:1–9.
9. Chadha MS, Comer JA, Lowe L, Rota PA, Rollin PE, Bellini WJ, et al. Nipah virus- associated encephalitis outbreak, Siliguri, India. *Emerg Infect Dis.* 2006;12(2):235–40. <https://doi.org/10.3201/eid1202.051247>
10. Krishnan S. Nipah outbreak in India and Bangladesh, WHO Communicable Disease Department Newsletter. 2007;4(2).
11. Daniels P, Ksiazek T, Eaton BT. Laboratory diagnosis of Nipah and Hendra virus infections. *Microbes Infect.* 2001;3:289–95. [https://doi.org/10.1016/S1286-4579\(01\)01382-X](https://doi.org/10.1016/S1286-4579(01)01382-X)
12. Giangaspero M. Nipah Virus. *Trop Med Surg* 2013;1:129.
13. Chong HT, Kamarulzaman A, Tan CT, Goh KJ, Thayaparan T, Kunjapan SR, et al. Treatment of acute Nipah encephalitis with ribavirin. *Ann Neurol.* 2001; 49(6):810–3. <https://doi.org/10.1002/ana.1062>
14. Bossart KN, Zhu Z, Middleton D, Klippel J, Crameri G, Bingham J, et al. A neutralizing human monoclonal antibody protects against lethal disease in a new ferret model of acute Nipah virus infection. *PLoS Pathog.* 2009; 5:e1000642. <https://doi.org/10.1371/journal.ppat.1000642>
15. Dawes BE, Kalveram B, Ikegami T, Juelich T, Smith JK, Zhanget L, et al. Favipiravir protects against Nipah virus infection in the hamster model. *Sci Rep.* 2018;8:7604. <https://doi.org/10.1038/s41598-018-25780-3>

Summary of the case

1	Patient (gender, age)	Male, 55
2	Final diagnosis	Nipah virus infection with cerebellar dysfunction
3	Symptoms	Fever, loose stools, vomiting, throat pain, gait ataxia
4	Medications	Ribavirin
5	Clinical procedure	-Antiviral Medication
6	Specialty	Internal Medicine, Neurology